

RESEARCH ARTICLE

Antioxidant and Anti-cancer effects of crude extracts from (*Vitis vinifera* L.) leaves on melanoma cells (SK-Mel and A375)

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ABSTRACT

This study aimed to estimate total phenol, the antioxidant and antiproliferative activities of crude extract derived from grape leaves (*Vitis vinifera* L.). Total phenol of grape leaves (*Vitis vinifera* L.) was extracted using aqueous and methanol solvent and dosed with Folin-Ciocalteu using spectrophotometry. The antioxidant activity was estimated by spectrophotometry in the existence of the DPPH radical, when the antiproliferative activity was evaluated by using MTT test on two melanoma cells (A375 and SK-MEL) as compared to the effect of Cisplatin. Results showed that extracts of grape leaves (*Vitis vinifera* L.) were rich in total phenols. Indeed, the extracts exhibited an antioxidant activity, when 1.98 mg/mL of methanol and aqueous crude extracts could inhibit 90.20 % and 77.78 % of DPPH* radical respectively. The proliferation of Melanoma cells A375 and SK-MEL is decreased with increasing concentration of water and methanolic extracts (1.136, 2.27, and 4.54 mg/mL) included in the culture throughout 72 h. Results offered that extract from grape leaves (*Vitis vinifera* L.) characterize with biological effect compounds such as phenols, with promise antioxidant activity, revealing an antiproliferative effect impact on A375 and SK-MEL malignant tumor cells compete with the synthetic molecule Cisplatin.

Keywords: *Vitis vinifera*; phenols; antioxidant; anti-melanoma; A375 and SK-MEL

INTRODUCTION

Oxidative stress is defined as an unbalance between the production of free radicals and reactive metabolites, called oxidants or reactive oxygen species (ROS), and their exclusion by protective mechanisms, named antioxidants (Reuter et al., 2010).

Antioxidants are defined as “any endogenous or exogenous substance which in low concentration relative to the oxidizable substrate prevents or slows the oxidation of that substrate” (Pastre and Priymenko, 2007). Antioxidants now appear as the keys to longevity and our allies to fight against modern diseases such cancer, diabetes and ageing. These are protective elements that act as free radical scavengers (Bartosz, 2003). There is now a renewed attention in phytochemicals as sources of bio-antioxidants. The purpose is to exploit them in foods

and pharmaceutical preparations to supplant synthetic antioxidants, which cause potential health risks due to their carcinogenic or mutagenic effects (Le Cren, 2012). Such as olive oil has been proposed by many experts as an important source of fat in the Mediterranean diet, it can form a fundamental influence on health benefits of Non-alcoholic fatty liver disease patients (Abenavoli et al., 2019).

Recently, the increased consumption of fruits and vegetables in its raw and processed form, or their various types of antioxidants containing as nutraceuticals, pharmaceuticals, and phytochemicals may decrease the risk of the progress of chronic human diseases (Jideani et al., 2020). Contrary following a recent study suggest a controlled consumption of citrus juice could limit the risk of skin cancer non-melanoma in postmenopausal women (Sakaki et al., 2021).

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Skin cancer refers to the three most common types of basal cell carcinoma (BCC) and squamous cell carcinoma (SCC) of the skin (collectively called non-melanoma skin cancer) and melanoma. (Yokoyama et al., 2011). Melanoma is the most invasive skin cancer, difficult to treat and potentially capable of metastasizing (Jaszewska et al., 2009). Since the mid-1960s, the incidence of melanoma has increased from 3 to 8 % each year in most people of European descent, with the largest increases in older men (Thompson et al., 2005).

Melanoma accounts for only 4 % of all skin cancers, but 80% of skin cancer deaths. When UV exposure is the primary cause of melanoma, only 14% of patients with metastatic melanoma survive for 5 years (Arlo et al., 2006). Direct mutagenicity on DNA, stimulates cellular components of the skin to produce growth factors, reduces the immune defenses of the skin, promotes melanin of reactive oxygen species (ROS), induces and prevents DNA damage, promotes malignant changes in the skin (Thompson et al., 2005).

Like the all cancers, genetic predispositions are identified in patients with a family history Avilés JA, (High and Lázaro 2006, Pho, 2006 and Robinson, 2007). Usually the first treatment used is surgical removal. Chemotherapy, radiation therapy and immunotherapy may also be used. New treatments like ipilimumab (Yervoy) are particularly effective against melanoma in advanced stage (Hodi et al., 2010).

Cisplatin complex is clinically used as adjunct tumor therapy to induce cancer cell death and tumor lesions through the induction of apoptosis (Florea et Büsselberg, 2011). Cisplatin has been clinically proven to fight several types of cancer, including sarcomas, soft tissue, bone, muscle, and vascular cancers. Despite the fact that such malignancies have recently gotten a better prognosis and treatment options (Dasari et Tchounwou, 2014).

Although plant extracts showed high anti-cancer and anti-proliferative activity (Chang et al., 2002; Nam et al.; 2003 and Lu et al., 2012). The isolation of the alkaloid from *Catharanthus roseus* (Apocynaceae) opened a new era in the use of plant materials as anticancer agents (Shoeb, 2006 and Manju et al., 2012). These compounds interfere with many aspects of tumor formation and progression in both *in vitro* and *in vivo* experiments. They induce cell cycle arrest in G1 or G2/M phase and inhibit the growth of various tumor cell lines by apoptosis (Lu et al., 2012).

Diarrhea and bleeding can be treated with grape leaves (Zargari, 1993). Grapes were also used to cure constipation, gastritis, enteritis, gout, and hemorrhagic diarrhea (Afzalzadeh et al., 2013). Different parts of grape vine have

been employed since antiquity because phenols are naturally present in many of these biological features.

DelCastilloAlonso et al., 2020 were able to identify 47 phenolic compounds in the skins and wines of Tempranillo grapevines exposed or non-exposed to close-to-ambient solar UV levels using appropriate filters, including flavonols, anthocyanins, flavanols, stilbenes, and hydroxycinnamic and hydroxybenzoic acids. In addition, 25 anthocyanins, 17 flavonols, 7 hydroxycinnamic acid derivatives, 2 stilbenes, and many flavan-3-ols have been found and measured in berry sections from two unique *Vitis vinifera* L. red grape genotypes (Moribel and Tinto Fragoso) (Pérez-Navarro et al., 2019).

The aims of this work are to estimate the total phenols extracted by aqueous and methanol from grape leaves (*Vitis vinifera* L.), to evaluate of their antioxidant potential by using the DPPH* test and antiproliferative activities by using MTT test on two melanoma cells (A375 and SK-MEL) as compared the effect of Cisplatin.

MATERIAL AND METHODS

Plant material

In August, mature leaves from the apical section of *Vitis vinifera* L. were harvested in Medea (warm temperate climate, average temperature 14.4 C°), Algeria. The leaves were cleaned under running water and dried at room temperature (25°C). Finally, they were ground into a fine powder and stored in a sterile bag under vacuum in the dark at 5 °C (Ferhi et al., 2019).

Extraction

Aqueous crude extract

The extraction is done as indicated (Sanogo *et al.*, 2006) with a few changes. It was performed at room temperature by macerating 40 g of the leaf-powder into one liter of sterile distilled water and stirring during 24 h. Then, the supernatant was filtered and frozen at – 30 °C.

Methanol crude extract

The methanol extract was performed as indicated (Mokale Kognou et al., 2011) with little changes. Ten gr of dried leaves were macerated in 50 mL of methanol/water 80 ° (v/v) during 24 h. Then, the solution was filtered with 0, 45 µm syringe Millipore filter and conserved at 4°C until needed.

Total phenolic (TP) content

The modified Folin-Ciocalteu technique was used to determine the total phenolic content (Ferhi et al., 2019).

1 mL of each extract was vortexed and centrifuged with 9 mL of ethanol (80%) (1:10 w/v). The Folin-Ciocalteu reagent (1mL) was then added to 200 μ L of each extract. A spectrophotometer was used to measure the absorbance at 760 nm (8453 Agilent Technologies, Santa Clara, CA, USA).

On the basis of a Gallic acid calibration curve (50 to 500 mg/L with $R^2 = 0.996$), the results were reported as milligrams of Gallic acid equivalent/g of dry weight.

Antioxidant activity

The scavenging activity was evaluated using the DPPH* test (Choi et al., 2002). The extracts' absorbance at 518 nm reduced as the concentration of extracts increased. The ability of the extracts to scavenge free radicals improved that the concentration of the extracts rose (Azad et al., 2015).

The test solution was prepared by adding 30 μ L of each extract at different concentrations (10, 20, 25, 50, 100, 200 mg/mL) to 3 mL of 0.3 mM DPPH methanol solution. However, to prepare the control solution, 3 mL of methanol was added to 30 μ L of the sample solution at all concentrations. Then, 3 mL of DPPH solution was added to 30 μ L for negative control. The blank for this solution is methanol. The solution was kept in the dark for 30 minutes at room temperature. The absorbance values were measured at 518 nm and converted to a percentage of antioxidant activity according to the following formula:

Antioxidant activity (%) = [(absorbance solution - absorbance control) \div absorbance control] \times 100

Cell culture

SK-MEL and A375 melanoma cell line preserved in the culture medium (RPMI) derives from The Roswell Park Memorial Institute 10 % fetal bovine serum (FBS) RPMI medium growing in T25 culture flask. Cultures were incubated at 37 °C in 5 % CO₂ incubator. While cells reach confluence need to be detached by trypsinization and to be "splitted", transferred to a new flask at a lower density and keep them growing.

MTT viability assay

In order to evaluate the antiproliferative activity of methanol and aqueous crude extracts of *Vitis vinifera* L. leaves on melanoma cells; a cell viability test is useful. The MTT assay is a colorimetric assay based on the ability of functional mitochondria of cells to reduce by succinate dehydrogenase enzyme (Mosmann, 1983). The MTT formazan crystals produced in detectable visible spectrometry after dissolution in DMSO (Diméthylsulfoxyde). Then, the amount of living cells surviving in the assessed cell culture is then used to determine the effects of the treatments studied on the overall growth of a given cell population.

Proliferation assay

The cells following (A375, SK-Mel) as used as 3×10^3 /well in 200 μ L of RPMI medium on a 96 well plate. Before, it needed to count cells by using (Talitm Image Based Cytometer Invitrogen Life Technology) and let them grow over night at 37 °C in a 5 % CO₂ incubator. After 24 hours substitute growth RPMI medium with new RPMI medium containing test compounds (methanol and aqueous crude extract) at final concentration of 4,54 mg/mL, 2,27 mg/mL and 1.136 mg/mL knowing that the mixture as follows 10 μ L of extract and 190 μ L of RPMI medium. The negative control is prepared only by adding RPMI medium but positive control is prepared by Cis-Platinum 10 μ M dilute in RPMI medium. It performed a test by replacing the extracts with methanol in the RPMI medium in order to see how methanol affected the two cell lines that were employed. Every test is performed three times. After one day, it substituted again RPMI medium with or without compounds and repeat the same treatment (treatment 2). Finally, after 48 hours since the first treatment, observed cells at the microscope and stopping viability with assay (MTT method) by adding 20 μ L of MTT solution, 5 mg/mL in Phosphate-buffered saline (PBS) to each well, Incubate at 37 °C in the 5 % CO₂ incubator for 3-4 hours and remove the RPMI medium by aspiration then added 100 μ L/well DMSO, Shaking for 10 min with a shaker (ThermomexerConfortEppendorf) at room temperature. Read absorbance in a Microtiter plate (Sunrise_basicTecan) reader at 570 nm. The relative percentage of growth cells (viability of cells) is calculated by contribution to the negative control.

Relative percentage viability = Absorbance of sample / Absorbance of negative control \times 100

Statistical analysis

Data were expressed as mean \pm standard deviation. ANOVA test with Duncan's multiple range testing approach was used for statistical analysis (DMRT). This study was conducted using the SPSS 25.0 Windows program. P values < 0.05 were considered significant.

RESULTS

Total phenolic content and DPPH Radical scavenging activity

Table 1 showed values of IC₅₀ and Total phenols (TP).

In comparison, the aqueous crude extract to the methanol crude extract had much more TP (approximately 1.5 times) ($P < 0.05$). The observed TP content discrepancy could be caused by the polarity of the methanol.

In the presence of the DPPH radical, the antioxidant activity of the leaf extracts was measured using spectrophotometry.

The methanol and aqueous crude extract of grape leaves (*Vitis vinifera*. L.) exhibited an antioxidant activity and able to trap the DPPH radical. The antioxidant capacity is presented by the IC₅₀ coefficient, it is the quantity of antioxidant required to reduce 50 % of the concentration of the free radical DPPH. The IC₅₀ is calculated from the regression line formed from percentages of inhibitions. For the calculation of these values, Microsoft Excel software was used.

The percentage of scavenging is related to the crude extract concentration ($R^2 > 0.90$).

The maximum trapping DPPH radical (90.20 %) is given by the methanol extract 1.98 mg/mL. The same concentration gave (77.78 %) with the aqueous crude extract. The values of IC₅₀ are inversely related to antioxidant capacity, the lower IC₅₀ means higher antioxidant capacity of compounds. The least effective extracts are those of higher IC₅₀ values. Indeed, with IC₅₀s of 0.76 mg/mL and 1.1 mg/mL, respectively, methanol extract had stronger free radical scavenging activity than aqueous extract ($P < 0.05$) (Table 1).

Effect of grape leaves (*Vitis vinifera*. L.) methanol and aqueous crude extracts on Cell Proliferation

The antiproliferative activity of grape leaves (*Vitis vinifera*. L.) using MTT test on melanoma cells (A375 and SK-MEL) are documented in Fig. (1, 2), Photos (1, 2).

The MTT assay assesses mitochondrial activity and was chosen as a biochemical test to evaluate cell viability. In this test, a Tétrazolium salts is used as a substrate. Dehydrogenase enzymes in active mitochondria cleave the Tétrazolium ring, conducted to the production of formazan.

Thus, the yellow substrate is transformed in a blue-colored product. The survival cells after treatment with methanolic and water extract appeared with blue and death cells appeared no-colored or like water.

(W1, W2, W3: cells treated with different doses of aqueous extract from highest to lowest dose, M1, M2, M3: cells treated with different doses of methanol extract from highest to lowest dose, Cis: cells treated with

Cisplatinium,OH: cells treated only with methanol, Blank: only medium).

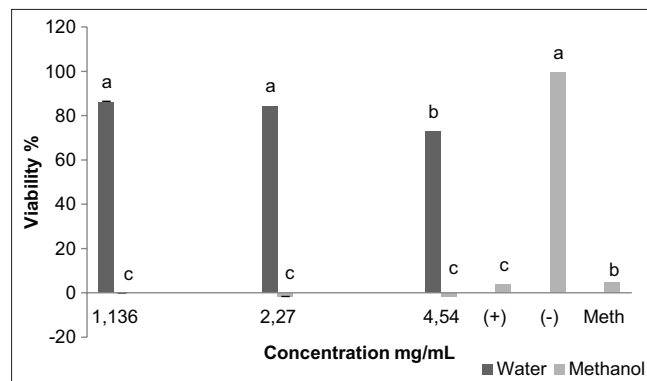


Fig 1. Effect of Methanolic and Aqueous crude extract on SK-Mel cell proliferation (untreated group: concentration = (-), group treated by Cisplatinium: (+), Methanol: (Meth)). Data are expressed as mean \pm SD, n = 3. According to Duncan's Multiple Range Test, bars with dissimilar letters within a group are substantially different at $P < 0.05$. (DMRT).

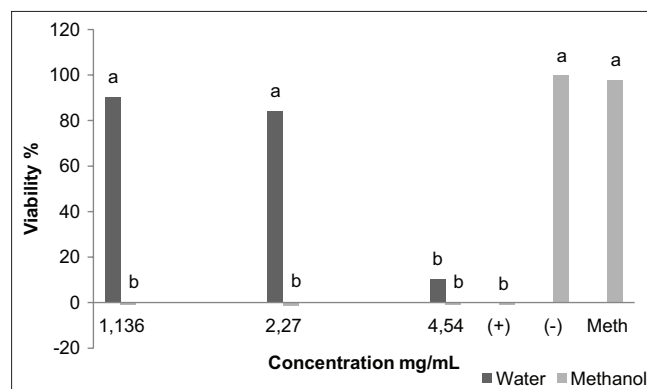


Fig 2. Effect of Methanolic and Aqueous crude extract on A375 cell proliferation (untreated group: concentration = (-), group treated by Cisplatinium: (+), Methanol: (Meth)). According to Duncan's Multiple Range Test, bars with dissimilar letters within a group are substantially different at $P < 0.05$. (DMRT).

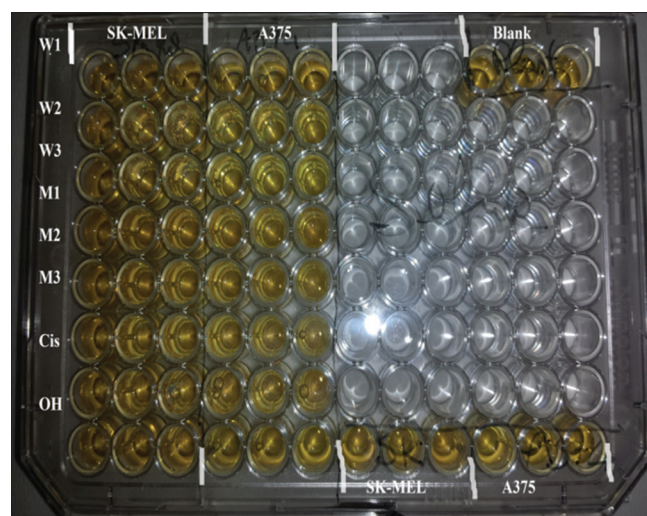


Photo 1. Plate with MTT before incubation

Table 1: IC₅₀ and TP of methanol and aqueous crude extract of grape leaves (*Vitis vinifera* L.)

Extracts	TP (mg GAE/g DW) \pm SD	IC ₅₀ (mg/mL) \pm SD	R ²
Methanol	23.0 \pm 21.3 ^a	0,76 \pm 0.08 ^a	0.991
Aqueous	16.35 \pm 2.06 ^b	1,10 \pm 0.12 ^b	0.984

TP: Total Phenols, GAE: Gallic acid equivalent; DW: Dry weight; SD: standard deviation; sample concentration at which 50% of the free radical activity was inhibited, R²: the correlation coefficient, The values with dissimilar letters are significantly different at $P < 0.05$

Effect of methanolic and aqueous extracts of grape leaves (*Vitis vinifera* L.) on SK-Mel Proliferation

SK-Mel cell viability was significantly reduced after incubation with methanol extract ($P = 0.001$) (cell proliferation is expressed as the average percentage of viable and untreated cells). (Photo 2). While, the aqueous extract was not toxic for SK-Mel cells, the cell proliferation reduced but insignificantly ($P > 0.05$); exceptionally the maximum dose (4.54 mg/mL) could inhibit significantly SK-Mel cell proliferation. The maximum growth inhibition was obtained using methanol extract (~100 %) followed by Cisplatinum (> 97 %), representing the positive control, and finally the aqueous extract 4.54 mg/mL (> 72 %). The methanol was toxic for SK-Mel cells and could inhibit significantly ($P < 0.05$) growth of but less than methanol extract. We can conclude that SK-Mel cells were sensitive to methanol (Photo 2).

Effect of methanolic and aqueous crude extracts of grape leaves (*Vitis vinifera* L.) on A375 cell Proliferation

Both crude extracts significantly ($P < 0.05$) decreased A375 cell multiplication, similar to what was seen in SK-Mel cells (Figure 2). Specifically, that methanol extract was more efficacious on cells than water extract exceptionally the highest dose of water extract (4.45 mg/mL), when its effect was powerful on A375 cell proliferation. Maximum growth inhibition rate was obtained by methanol extract (100 %), followed by Cisplatinum and the highest dose of water extracts (Figure 2). The A375 cells showed more resistant to methanol than SK-Mel cells also it was not toxic on A375 cells. In this case, it can be deduced that the methanol extract of grape leaves (*Vitis vinifera* L.) had a similar or better effect on the growth of A375 cells than that of cisplatin.

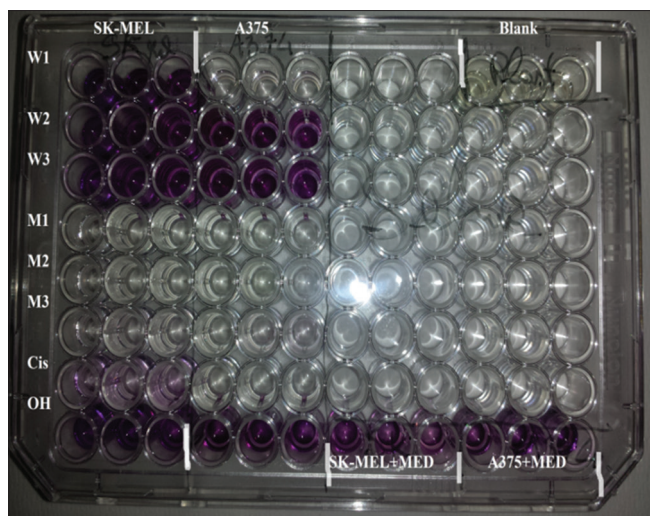


Photo 2. Plate with MTT after incubation

DISCUSSION

Antioxidants now appear as the keys to longevity and our allies to fight against modern diseases. These are protective elements that act as free radical scavengers (Bartosz, 2003). The interesting activity of grape leaves (*Vitis vinifera* L.) crude extract is explained by the presence of phenolics compounds, these results are accorded to those obtained previously by Nassr-Allah et al, 2009, Popovici et al, 2009, Ba et al, 2010, Zheng et al., 2010 and Katalinic et al., (2009 and 2013). These discoveries have given interest in grape leaves (*Vitis vinifera* L.) as a possible source of chemicals as nutritional and biological value. When, Monagas et al., 2006^a and Ferhi et al., 2019¹ found that grape leaves (*Vitis vinifera* L.) contained high level of phenols; anthocyanins, flavonols and trans-caftaric acid.

The anticancer activities of essential oils, aqueous and organic solvent extracts of many aromatic plants were evaluated *in vivo* and *in vitro*, *Luffa aegyptiaca* (sponge gourd), *Cassia italica* (Senegal senna), *Ocimum basilicum* (basil), *Colocasia antiquorum* (taro), *Beta vulgaris* (beet), fruit of *Capsicum frutescens* (chili pepper) and fruit, leaves, root of *Morinda citrifolia* (Brown, 2012). Also, the extracts from *Urtica membranacea* (Urticaceae), *Artemisia monosperma* (Asteraceae), and *Origanum dayi* post (Labiatae) demonstrated toxicity in a variety of human tumor cell lines and primary cultures derived from patient biopsies. (Solowey et al., 2014).

Similarly, the methanol extracts of *Dendrosicyos cotranra*, *Withania aduensis*, *Withania eibeckii*, *Dracenacinnabari* and *Buxus hildebrandtii* showed a high toxicity on all tumor cell lines with IC_{50} values ranging between 0.29 and 5.54 mg/mL (Mothana et al., 2007). In all cell lines, *Scutellaria baicalensis* exhibited a significant dose-dependent growth reduction, with IC_{50} values of 1.1, 0.9, 0.52, 0.82, 1.1, 1.5, 1.0, and 1.2 mg/mL on HepG2, MCF-7, PC-3, LNCaP, KM-12, HCT-15, KB, and SCC-25 cells, respectively (Ye et al., 2002). Other plants like *Inula graveolens*, *Salvia dominica*, *Conyza canadensis* and *Achillea santolina* showed also showed strong antiproliferative activity (Abu-Dahab and Affi, 2007).

In addition, at IC_{50} of 30 Mm (9.5 μ g/mL), the phenolic compounds extracted from *Rabdosia japonica* proved cytotoxicity against the murine B16-F10 melanoma cell line (Nitoda et al., 2008). The aqueous extracts of aerial part of *Tetraena gaetula* and root bark of *Berberis hispanica* (at 80 and 300 mg/kg, respectively) had a strong antiproliferative activity (El Youbi et al., 2012).

habitually, thymol is used as an inhibitor of the enzymatic system of melanin production, otherwise, it proved moderate cytotoxicity with an IC_{50} value of 400 μ M but

no antimelanogenic action on B16-F10 melanoma cells (60.09 µg/mL) (Satooka and Kubo, 2012).

Inhibition of 72.58 % of melanoma metastatic colony formation in the lungs was obtained by treating animals with Withania extract at 20 mg/dose/animal, while intraperitoneally injection of 500 g/dose/animal of Withanolide D inhibited 69.84 percent of B16F-10 development (Leyon and Kuttan, 2004).

Another study found that Triterpene Extract from Mistletoe, with an IC₅₀ of 2.675 mg/mL, suppressed the growth of B16.F10 cells in a dose-dependent manner (Struh et al., 2012).

The 200 µg/mL of 1,1-dimethylallyl caffeate (DMAC), 3-methyl-3-butenyl caffeate, pinocembrin, benzyl ferulate, benzyl isoferulate and tectochrysin provided excellent antiproliferative activity against all human gastrointestinal cancer cell lines (> 80%) (DLD-1 colon adenocarcinoma, HCT-116 colon carcinoma, KYSE-30 oesophageal squamous cancer, and NCI-N87 gastric carcinoma); while Pinobanksin-3-O-acetate, 5-phenyl-Penta-2,4-dienoic acid, and pinostrobinchalcone showed moderate to high activity against all cell lines except KYSE30, however, p-coumaric acid was inactive Catchpole et al., (2015).

Finally, Ferhi et al., (2019) provide that the extract of grape leaves (*Vitis vinifera* L.) grown in (Algeria) showed power antiproliferative effect on MCF-7 breast cancer cells and HepG2 hepatocarcinoma cells.

CONCLUSION

For the first time, it discovered in this research that the methanolic and aqueous crude extracts of grape leaves (*Vitis vinifera*) grown in Medea area in Algeria promoted an interesting source of not dangerous bioactive molecules like phenols who provided antioxidant and anti-melanoma activities, these can even conquer the effect of Cisplatin. So, it suggested that these extracts might use as available source of bio-antioxidants and as matrix for preparing medicaments countering cancer cells proliferation.

COMPLIANCE WITH ETHICAL STANDARDS

Conflict of interest

There are no conflicts of interest declared by the authors.

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AUTHORS' CONTRIBUTIONS

S.F. is the leading author, who developed the idea, practical works, wrote and revised the article. T.C. helped to discuss the results, S.G. helped to find journal and submitted the work, R.M.C and G.D. provided technical guidance during simulations and experiments, the necessary technical tools for the realization of this work S.Z. is the supervisor of this work. G.D. supervised the work and is the coordinators of the guesting Institute

REFERENCES

- Abenavoli, L., M. Milanović, N. Milić, F. Luzzza and A. M. Giuffrè. 2019. Olive oil antioxidants and non-alcoholic fatty liver disease. *Exp. Rev. Gastroenterol. Hepatol.* 13: 739-749.
- Abu-Dahab, R. and F. Afifi. 2007. Antiproliferative activity of selected medicinal plants of Jordan against a breast adenocarcinoma cell line (MCF7). *Sci. Pharm.* 75: 121-146.
- Afzalzadeh, M. R., A. A. Papahn, A. Amirzargar, M. K. Varnamkhasti, H. Ganjali and E. G. Mombeni. 2013. Effect of *Vitis vinifera* leave hydro-alcoholic extract on reproductive parameters in adult normal male rats. *J. Phys. Pharm. Adv.* 6: 159-166.
- Arlo, J. M. and M. C. Jr. Mihm. 2006. Melanoma. *N. Engl. J. Med.* 355: 51-65.
- Avilés, J. A. and P. Lázaro. 2006. Genetic predisposition in cutaneous melanoma. *Actas Derm. Sifiliograficas.* 97: 229-240.
- Azad, A. N., V. Hakimzadeh and E. Golmakani. 2015. Phenolic contents and antioxidants activity from aerial parts of *Phlomis herba-venti* L. subsp. kopetdaghensis. *J. Appl. Environ. Biol. Sci.* 4: 54-58.
- Ba, K., E. Tine, J. Destain, N. Cissé and P. Thonart. 2010. Étude comparative des composés phénoliques, du pouvoir antioxydant de différentes variétés de sorgho sénégalais et des enzymes amylolytiques de leur malt. *Biotechnol. Agron. Soc. Environ.* 14: 131-139.
- Bartosz, G. 2003. Generation of reactive oxygen species in biological systems. *Comment Toxicol.* 9: 5-21.
- Brown, A. C. 2012. Anticancer activity of *Morinda citrifolia* (Noni) fruit: A review. *Phytother. Res.* 26:1427-1440.
- Catchpole, O., K. Mitchell, S. Bloor, P. Davis and A. Suddes. 2015. Antiproliferative activity of New Zealand propolis and phenolic compounds vs human colorectal adenocarcinoma cells. *Fitoterapia.* 106:167-174.
- Chang, H. C., W. C. Hung, M. S. Huang and H. K. Hsu. 2002. Extract from the leaves of *Toona sinensis* roemor exerts potent antiproliferative effect on human lung cancer cells. *Am. J. Chin. Med.* 30: 307-314.
- Choi, C. W., S. C. Kim, S. S. Hwang, B. K. Choi, H. J. Ahn, M. Y. Lee and S. Kim. 2002. Antioxidant activity and free radical scavenging capacity between Korean medicinal plants and flavonoids by assay-guided comparison. *Plant Sci.* 163: 1161-1168.

- Dasari, S. and P. B. Tchounwou. 2014. Cisplatin in cancer therapy: Molecular mechanisms of action. *Eur. J. Pharmacol.* 740: 364-378.
- Del-Castillo-Alonso, M. Á., L. Monforte, R. Tomás-Las-Heras, J. Martínez-Abaigar and E. Núñez-Olivera. 2020. Phenolic characteristics acquired by berry skins of *Vitis vinifera* cv. Tempranillo in response to close-to-ambient solar ultraviolet radiation are mostly reflected in the resulting wines. *J. Sci. Food Agric.* 100: 401-409.
- El Youbi, A. E. H., D. Bousta, B. Jamoussi, H. Greche, L. El Mansouri, J. Benjilali and S. H. Soidrou. 2012. Activités antioxydante, apoptotique et antiproliférative de *Tetraena gaetula* (Emb. & Maire) Beier & Thulin et de *Berberis hispanica* Boiss. & Reut. originaires du Maroc. *Phytothérapie.* 10: 151-160.
- Ferhi, S., Santaniello, S., S. Zerizer, S. Cruciani, A. Fadda, D. Sanna, A. Dore, M. Maioli and G. D'hallewin. 2019. Total phenols from grape leaves counteract cell proliferation and modulate apoptosis-related gene expression in MCF-7 and HepG2 human cancer cell lines. *Molecules.* 24: 612.
- Ferhi, S., S. Zerizer and G. D'hallewin. 2019. The effects of grape leaves extract on hyperhomocysteinemia induced inflammatory endothelial damage in cardiovascular diseases. *Appl. Ecol. Environ. Res.* 17: 1989-2003.
- Florea, A. M. and D. Büsselberg. 2011. Cisplatin as an anti-tumor drug: cellular mechanisms of activity, drug resistance and induced side effects. *Cancers.* 3: 1351-1371.
- High, W. A. and W. A. Robinson. 2007. Genetic mutations involved in melanoma: A summary of our current understanding. *Adv. Dermatol.* 23: 61-79.
- Hodi, F. S., S. J. O'Day, D. F. McDermott, R. W. Weber, J. A. Sosman, J. B. Haanen and W. J. Urba. 2010. Improved survival with ipilimumab in patients with metastatic melanoma. *N. Engl. J. Med.* 363: 711-723.
- Jaszewska, E., A. Kosmider, A. K. Kiss and M. Naruszewicz. 2009. Pro-oxidative and pro-apoptotic action of defatted seeds of *Oenothera paradoxa* on human skin melanoma cells. *J. Agric. Food Chem.* 57: 8282-8289.
- Jideani, A. I., H. Silungwe, T. Takalani, A. O. Omolola, H. O. Udeh and T. A. Anyasi. 2021. Antioxidant-rich natural fruit and vegetable products and human health. *Int. J. Food Properties.* 24: 41-67.
- Katalinic, V., I. Generalić, D. Skroza, I. Ljubenkov, A. Teskera, I. Konta and M. Boban. 2009. Insight in the phenolic composition and antioxidative properties of *Vitis vinifera* leaves extracts. *Croat. J. Food Sci. Technol.* 1: 7-15.
- Katalinic, V., S. S. Mozina, I. Generalic, D. Skroza, I. Ljubenkov and A. Klancnik. 2013. Phenolic profile, antioxidant capacity, and antimicrobial activity of leaf extracts from six *Vitis vinifera* L. varieties. *Int. J. Food Properties.* 16: 45-60.
- Le Cren, F. 2012. Les Antioxydants: La révolution du XXI^e siècle. Les Éditions Québec-livres.
- Leyon, P. V. and G. Kuttan. 2004. Effect of *Withania somnifera* on B16F-10 melanoma induced metastasis in mice. *Phytother. Res.* 18: 118-122.
- Lu, J. J., J. L. Bao, X. P. Chen, M. Huang and Y. T. Wang. 2012. Alkaloids isolated from natural herbs as the anticancer agents. *Evid. Based Complement. Altern. Med.* 2012:485042.
- Manju, K., R. K. Jat and G. Anju. 2012. A review on medicinal plants used as a source of anticancer agents. *Int. J. Drug Res. Technol.* 2: 177-183.
- Mokale Kognou, A. L., R. A. N. Ngane, J. R. Kuate, M. L. K. Mogtomo, A. T. Tiabou, R. S. Mouokeu and P. H. A. Zollo. 2011. Antibacterial and antioxidant properties of the methanolic extract of the stem bark of *Pteleopsis hylodendron* (Combretaceae). *Chemother. Res. Pract.* 2011: 218750.
- Monagas, M., B. Hernández-Ledesma, C. Gómez-Cordovés and B. Bartolomé. 2006a. Commercial dietary ingredients from *Vitis vinifera* L. leaves and grape skins: Antioxidant and chemical characterization. *J. Agric. Food Chem.* 54: 319-327.
- Mosmann, T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Methods.* 65: 55-63.
- Mothana, R. A. A., R. Gruenert, U. Lindequist and P. J. Bednarski. 2007. Study of the anticancer potential of Yemeni plants used in folk medicine. *Pharm. Int. J. Pharm. Sci.* 62: 305-307.
- Nam, N. H., C. W. Lee, D. H. Hong, H. M. Kim, K. H. Bae and B. Z. Ahn. 2003. Antiinvasive, antiangiogenic and antitumour activity of *Ephedra sinica* extract. *Phytother. Res.* 17: 70-76.
- Nassr-Allah, A. A., A. M. Aboul-Enein, K. M. Aboul-Enein, D. A. Lightfoot, A. Cocchetto and H. A. El-Shemy. 2009. Anti-cancer and anti-oxidant activity of some Egyptian medicinal plants. *J. Med. Plants Res.* 3: 799-808.
- Nitoda, T., T. Isobe and I. Kubo. 2008. Effects of phenolic compounds isolated from *Rabdosia japonica* on B16-F10 melanoma cells. *Phytother. Res.* 22: 867-872.
- Pastre, J. and N. Priymenko. 2007. Intérêt des anti-oxydants dans l'alimentation des carnivores domestiques. *Rev. Méd. Vét.* 1: 180-189.
- Pérez-Navarro, J., P. M. Izquierdo-Cañas, A. Mena-Morales, J. Martínez-Gascuña, J. L. Chacón-Vozmediano, E. García-Romero and S. Gómez-Alonso. 2019. Phenolic compounds profile of different berry parts from novel *Vitis vinifera* L. red grape genotypes and Tempranillo using HPLC-DAD-ESI-MS/MS: A varietal differentiation tool. *Food Chem.* 295: 350-360.
- Pho, L., D. Grossman and S. A. Leachman. 2006. Melanoma genetics: A review of genetic factors and clinical phenotypes in familial melanoma. *Curr. Opin. Oncol.* 18: 173-179.
- Popovici, C., I. Saykova and B. Tylkowski. 2009. Evaluation de l'activité antioxydante des composés phénoliques par la réactivité avec le radical libre DPPH. Université technique de Moldova, Kishinev, Moldavie; Université de technologie chimique et de métallurgie, Sofia, Bulgarie. *Rev. Gén. Ind.* 4: 25-39.
- Reuter, S., S. C. Gupta, M. M. Chaturvedi and B. B. Aggarwal. 2010. Oxidative stress, inflammation, and cancer: How are they linked? *Free Radic. Biol. Med.* 49: 1603-1616.
- Sakaki, J. R., M. M. Melough, M. B. Roberts, C. B. Eaton, A. H. Shadyab, A. A. Qureshi and E. Cho. 2021. Citrus consumption and the risk of non-melanoma skin cancer in the women's health initiative. *Cancers.* 13: 2173.
- Sanogo, R., D. Diallo, S. Diarra, C. Ekoumou and F. Bougoudogo. 2006. Activité Antibactérienne et Antalgique de Deux Recettes Traditionnelles Utilisées Dans le Traitement des Infections Urinaires et la Cystite au Mali. *Médecine d'Afrique Noire, Traitement traditionnel Mali Médical*, pp. 19-24.
- Satooka, H. and I. Kubo. 2012. Effects of thymol on B16-F10 melanoma cells. *J. Agric. Food Chem.* 60: 2746-2752.
- Shoeb, M. 2006. Anti-cancer agents from medicinal plants. *Bangladesh J. Pharmacol.* 1: 35-41.
- Solowey, E., M. Lichtenstein, S. Sallon, H. Paavilainen, E. Solowey and H. Lorberboum-Galski. 2014. Evaluating medicinal plants for anticancer activity. *Sci. World J.* 2014: 721402.
- Struh, C. M., S. Jäger, C. M. Schempp, A. Scheffler and S. F. Martin. 2012. A novel triterpene extract from mistletoe induces rapid apoptosis in murine B16. F10 melanoma cells. *Phytother. Res.* 26: 1507-1512.

- Thompson, J. F., R. A. Scolyer and R. F. Kefford. 2005. Cutaneous melanoma. *Lancet*. 365: 687-701.
- Ye, F., L. Xui, J. Yi, W. Zhang and D. Y. Zhang. 2002. Anticancer activity of *Scutellaria baicalensis* and its potential mechanism. *J. Altern. Complement. Med.* 8: 567-572.
- Yokoyama, S., S. L. Woods, G. M. Boyle, G. L. Aoude, S. MacGregor, V. Zismann and K. M. Brown. 2011. A novel recurrent mutation in MITF predisposes to familial and sporadic melanoma. *Nature*. 480: 99-103.
- Zargari, A. 1993. *Medicinal Plants*. Vol. 1. Tehran University Publications, Tehran, pp. 5-362.
- Zheng, J., H. Li, C. Ding, Y. Suo, L. Wang and H. Wang. 2011. Anthocyanins composition and antioxidant activity of two major wild *Nitraria tangutorun* Bobr. variations from Qinghai-Tibet Plateau. *Food Res. Int.* 44: 2041-2046.