Effect of inland salt-affected soil on physiological and growth characters of Eucalypt tree (*Eucalyptus camaldulensis* Dehnh.)

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Abstract: *Eucalyptus* tree is a high potential plant species as a deep root system for water absorption to prevent the salt rise-up, a large bushy to cover the surface area and to detoxify the toxic salt ions. The aim of this research is to investigate the responses of *Eucalyptus camaldulensis* to different sites of inland salinity. Survival percentage of salt-stressed eucalypt was decreased, relating to salt contamination and negatively correlated with proline accumulation. Water use efficiency in the salt-stressed leaves was negatively related to electro conductivity (EC) values in the salt-affected soil. Chlorophyll a (Chl_a) and total chlorophyll (TC) degradations were positively related to maximum quantum yield of photosystem II (PSII) (F_v/F_m) (r^2 =0.89) and quantum efficiency of PSII (Φ_{PSII}) (r^2 =0.95), leading to low net photosynthetic rate (P_n) (P_n) (P_n) and to reduce the growth characteristics in term of plant height and circumstance. The survived eucalypt plants in the severe salinity land containing high salt contamination (1.7-2.0% salt) may be further applied as potential plant species to remediate the inland salinity soil.

Keywords: chlorophyll *a* fluorescence, electro conductivity, eucalyptus, growth, net-photosynthetic rate, photosynthetic pigments, salinity

تأثير التربة بالمناطق الداخلية المتأثرة بالملوحة على وظائف الفسيولويجة وخصائص النمو لنبات الكينا Eucalyptus camaldulensis Dehnh

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الملخص: يعتر نبات الكينا من الأنواع النباتية التي تمتلك النظام الجذري العميق لامتصاص المياه والحيلولة دون ارتفاع نسبة الملوحة وتميز ها بالمجوع الخضري وتغطية مساحات سطحية وإزالة ايونات الملح السامة . الهدف من هذه الدراسة هو التحقق من مدى استجابة نبات (Eucalyptus camaldulensis Dehnh) في مواقع متعددة من المناطق الداخلية المتأثرة بالملوحة نسبة البقاء لشجرة الكينا المتأثرة بالملوحة شهدت انخفاضا كبير ايرجع ذلك لتجمع الملوحة والعلاقة السلبية مع تراكم عنصر البرولين . نسبة كفاءة استخدام المياه في أور اق الأشجار المتأثرة بالملوحة شهدت اتجاها سلبا نتيجة لقيم التوصيل الكهربائي للقيم التوصيلية الكهربائية . في التربة المتأثرة بالملوحة وتبين أن عنصر الكلورفيل (A) انخفض لارتباطه الايجابي من الحد الأقصى من العائد من البناء الضوئي الثاني (PSII (Φ_{PSII}) الخفاض في معدل التمثيل الضوئي الثاني المباث وما يتصل ذلك من ظروف النمو المختلفة . ووجد أيضا أن نبات الكينا المقاوم في أراضى شديدة الملوحة والتي تحتوى على ملوثات (Φ_{PSII}) . يمكن أن يتم تطبيقها مستقبلا كنباتات لها قدرة على إصلاح التربة في المناطق الداخلية المتأثرة بالملوحة العالية.

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Introduction

Salt affected soil is one of environmental issues to become a serious problem, inhibiting plant growth and development prior to loss of productivity, especially in crop species being defined as glycophyte (Hasegawa et al., 2000; Qadir et al., 2008). Salt affected soils (EC > 4 dS m⁻¹) can be characterized by the excess levels of water-soluble salts, including sodium chloride (NaCl), sodium sulfate (Na₂SO₄), calcium chloride (CaCl₂) and magnesium chloride (MgCl₂), among others. Those salts are easily water soluble and played as a positive charge i.e. Na⁺, Ca²⁺, Mg²⁺, and K⁺, and a negative charge i.e. Cl^2 , SO_4^{2-} , HCO^3 , CO_3^{2-} , and NO_3^{-} (Tanji, 2002). In inland salinity, NaCl is a dominant salt contamination in the soil. It is a small molecule when oxidized by water, producing sodium ions (Na⁺) and chloride ions (Cl-), which are easily absorbed by the root tissues of higher plants and transferred to the whole plant using xvlem uploading channels (Maathuis Amtmann, 1999; Tester and Davenport, 2003; Rodriguez-Navarro and Rubio, 2006). In susceptible plant species, ionic and osmotic stresses derived from salt ions at the cellular level have been evidently identified (Mansour and Salama, 2004; Chinnusamy et al., 2005). There are many procedures that can be used to improve salt affected land, such as, water remediation leaching, chemical phytoremediation (Ahmad and Chang, 2002; Sharma and Minhas, 2005; Qadir et al., 2007). A phytoremediation technique is an effective way to solve the saltaffected soil problem (Turner and Ward, 2002; Qadir and Oster, 2004; Qadir et al., 2007; Jianfeng et al., 2008; Kiliç et al., 2008).

Eucalyptus camaldulensis is an original species from Australia and widely distributed in many regions of the world with a short rotation (Rockwood et al., 2008) for pulp and paper production. It is a miracle plant to be grown in the drought and saline soil conditions (Madsen and

Mulligan, 2006; Dale and Dieters, 2007). The objective of present study was to investigate the adaptabilities including survival percentage, proline accumulation, photosynthetic abilities and growth performances as indicator of eucalypt tree in the salt-affected soil.

Materials and methods

Plant materials

Eucalyptus plantlets were propagated and photo-autotrophically acclimatized, according to Cha-um et al. (2003). The plantlets were directly transplanted to open plastic bags (size 10W×10L×20H cm), containing a mixture of two parts of soil and one part of vermiculite. transplanted plantlets were incubated in a glasshouse at 30±2°C 75±5% relative humidity (RH) and 500±100 µmol m⁻² s⁻¹ photosynthetic proton flux density (PPFD) by natural light intensity at plant level with hd⁻¹ photoperiod. Six-month-old 10 transplanted plantlets were acclimatized by irrigation with 50 mM NaCl and 50% shading natural light intensity for 7 days and then directly planted in salinity field trials, plot site I, II and III at Ban Muang (17°5'0"N 103°35'0"E) Sakol Nakron, Northeastern region of Thailand different salt levels, 0.0-0.1% (control) and/or 1.7-2.0% salt (291-342 mM NaCl) (Table 1). Survival percentage, proline content, chlorophyll content, chlorophyll a fluorescence, photosynthetic abilities and growth performances were measured after planted in the field trial for 10 months.

The proline content of the leaves was extracted and analyzed according to the method of Bates et al. (1973). Fifty-milligrams of leaf materials were ground in liquid nitrogen using mortar. The homogenate powder was mixed with 1 mL aqueous sulfosalicylic acid (3% w/v) and filtered through filter paper (Whatman #1, England). The extracted solution was reacted with an equal volume of glacial acetic acid and ninhydrin reagent (1.25 mg ninhydrin in 30 mL glacial acetic acid and 20 mL 6 M H₃PO₄) and incubated at 95°C

for 1 h. The reaction was terminated by placing the container in an ice bath. The reaction mixture was mixed vigorously with 2 mL toluene. After cooling to 25°C,

the chromophore was measured by spectrophotometer DR/4000 at 520 nm using L-proline as a standard.

Table 1. Salt concentration, electro conductivity (EC) and soil pH in salt-affected sites in Northern region of Thailand.

Plot site	Salt (%)	EC (dS m ⁻¹)	рН	
I	0.1	2.1	5.87	
	1.7	13.3	4.83	
II	0.0	1.4	5.49	
	1.8	13.4	4.53	
III	0.1	3.7	5.58	
	2.0	16.1	4.72	

Plot sites I, II and III are located on 17°5′0″N 103°35′0″E to produce sodium chloride as salt mining using sun dry technique.

Chlorophyll a (Chla), chlorophyll b and total chlorophyll concentrations were determined following the method of Shabala et al. (1998) and the total carotenoids (C_{x+c}) concentration was determined following the method of One Lichtenthaler (1987).hundred milligrams of leaf material was collected and placed in a 25 mL glass vial, along with 10 mL 95.5% acetone, and blended using a homogenizer. The glass vials were sealed with parafilm to prevent evaporation, and stored at 4°C for 48 h. Chl_a and Chl_b concentrations were measured using a UV-visible spectrophotometer at 662 nm and 644 nm wavelengths. The C_{x+c} concentration was also measured by spectrophotometer at 470 nm.

Chlorophyll *a* fluorescence emission from the adaxial surface on the third leaf from the shoot tip was monitored using a Fluorescence Monitoring System (FMS 2; Hansatech Instruments Ltd., Norfolk, UK) in the pulse amplitude modulation mode, as previously described by Loggini et al. (1999). A leaf, adapted to dark conditions for 30 min using leaf-clips, was initially exposed to the modulated measuring beam of far-red light (LED source with typical

peak at wavelength 735 nm). Original (F₀) and maximum (F_m) fluorescence yields were measured under weak modulated red light ($<0.5\mu$ mol m⁻² s⁻¹) with 1.6 s pulses of saturating light (>6.8 µmol m⁻² s⁻¹ PAR), and calculated using FMS software for Windows®. The variable fluorescence vield (F_v) was calculated by the equation, F_m-F_0 . The ratio of variable to maximum fluorescence (F_v/F_m) was calculated as maximum quantum yield of photochemistry. The photon yield of PSII (Φ_{PSII}) in the light was calculated by Φ_{PSII} = $(F_m'-F)/F_m'$ after 45 s illumination, when steady state was achieved.

Net photosynthetic rate (P_n) , transpiration rate (E; mmol m⁻² s⁻¹) and stomatal conductance (g_s; mol H₂O m⁻² s⁻¹) measured using a were Portable Photosynthesis System with an Infra-red Gas Analyser (IRGA; Model LI 6400, LI-COR® Inc, Lincoln, Nebraska, USA). E and g_s were measured continuously by monitoring the H₂O content of the air entering and existing in the IRGA headspace chamber. Water use efficiency (WUE) of acclimatized plantlets was calculated by the ratio of P_n to E (Cha-um et al., 2007).

The experiment was arranged in a Completely Randomized Design (CRD) with 10 replications (n=10). Analysis of variance (*ANOVA*) was computed using the SPSS software (SPSS for Windows, SPSS Inc., USA), as well as means in each treatment were compared by the Least Significant Difference (LSD).

Results and discussion

Survival percentage of eucalypt tree in salt-affected soil was significantly dropped to 30, 16 and 14%, relating to 1.7, 1.8 and 2.0% salt concentrations, respectively (Figure 1). In contrast, proline contents in the leaf tissues of eucalypt were enriched and peaked at 4.1 µmol g⁻¹ FW in plant grown under 2.0% salt contamination in plot site III (Figure 1). The electro conductivity (EC) in the soil solution was negatively related to water use efficiency (Figure 2A). An increasing salt concentration in the soil solution directly iniured the chlorophyll a (Chl_a), chlorophyll b (Chl_b), total chlorophyll (TC) and total carotenoids (C_{x+c}) contents (Table 2). As example,

photosynthetic pigments, Chl_a, Chl_b, TC and C_{x+c} were significantly dropped when exposed to 1.70% salt conditions in plot site I for 54.79, 53.75, 54.50 and 50.30%, respectively. The Chl_a and TC contents in the salt-stressed leaves was positively correlated with maximum quantum yield of PSII (F_v/F_m) (Fig. 2B) and quantum efficiency of PSII (Φ_{PSII}) (Figure 2 C) respectively, causing to low photosynthetic rate (P_n) (Figure 2D). The F_v/F_m , Φ_{PSII} , P_n and stomatal conductance (g_s) in the leaf tissues was significantly decreased by salt stress, whereas the different plot sites were unaffected (Table 3). In contrast, the transpiration rate (g_s) in salt stressed leaves was enhanced. Diminishing of photosynthetic abilities including F_v/F_m , Φ_{PSII} , P_n and E was depended on salt contamination in the soil and higher than those in control condition of all plot sites, leading to overall growth reduction. In the salt affected soil, growth performances including plant height and branch circumstance were significantly decreased (Figure 3 and Figure 4).

Table 2. Chlorophyll a (Chl_a), chlorophyll b (Chl_b), total chlorophyll (TC) and total carotenoids (C_{x+c}) contents in the leaf tissues of eucalypt tree cultivated in different salt-affected sites for 10 months.

Plot site	Salt	Chla	Chlb	TC	C_{x+c}
	(%)	(μg g ⁻¹ FW)			
I	0.1	139.8a	54.7a	194.5a	50.9a
	1.7	63.2c	25.3b	88.5c	25.3c
II	0.0	147.3a	49.8a	197.1a	53.1a
	1.8	84.8b	31.7b	116.5b	33.4b
Ш	0.0	133.7a	49.3a	183.0a	48.3a
	2.0	72.4bc	26.7b	99.1bc	29.8bc
Significant lev	el				
Plot site		**	NS	**	**
Salt		**	**	**	**
Plot site×Salt		**	*	**	**

Different letters in each column show significant difference at $p \le 0.01$ (**) by Least Significant Difference (LSD). NS and * are represented non significant and significant at $p \le 0.05$, respectively.

Survival percentage and physiological adaptabilities of eucalypt tree in the salt-affected soil were reduced relating to salt concentrations in the soil. Similarly, the survival percentage of eucalypt (*E.*

camaldulensis) in the salt-affected soil is reduced to 87.6, 72.4 and 70.5%, depending on cultivation periods, 3, 6 and 9 months, respectively (Shirazi et al., 2006). In addition, the survival percentage

of *E. camaldulensis* cultivated in the 300-400 mM NaCl is better than other species, including *E. populnea* and *E. citriodora* (Madsen and Mulligan, 2006). Proline content is enriched in *E. camaldulensis*

treated with 100 mM NaCl, relating to salt concentrations (50 and 100 mM NaCl) and salt exposure times (Woodward and Bennett, 2005).

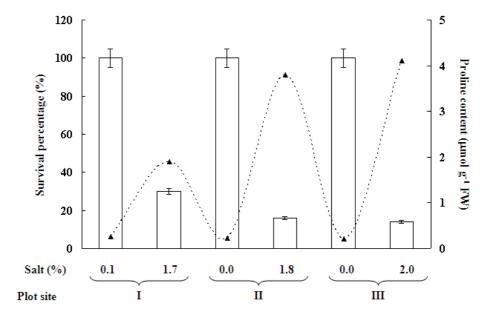


Figure 1. Survival percentage (white bar) and proline contents (dot line) in the leaf tissues of eucalypt tree cultivated in different salt-affected sites for 10 months.

Table 3. Maximum quantum yield of PSII (F_v/F_m) , photon yield of PSII (Φ_{PSII}) , net photosynthetic rate (P_n) stomatal conductance (g_s) and transpiration rate (E) in the leaf tissues of eucalypt tree cultivated in different salt-affected sites for 10 months.

Plot site	Salt	F_v/F_m	$\Phi_{ ext{PSII}}$	P _n	$\mathbf{g}_{\mathbf{s}}$	E
	(%)			$(\mu mol \ m^{-2}s^{-1})$	$(mmol m^{-2}s^{-1})$	(mmol m ⁻² s ⁻¹)
I	0.1	0.834a	0.785a	8.53b	47.2a	1.1c
	1.7	0.757b	0.633b	4.32c	17.6b	2.9a
II	0.0	0.870a	0.755a	10.49a	41.5a	1.0c
	1.8	0.751b	0.665b	4.20c	13.7b	2.5a
III	0.0	0.869a	0.786a	8.81b	36.6a	0.9c
	2.0	0.761b	0.644b	3.36c	14.5b	1.7b
Significan	t level					_
Plot site		NS	NS	**	**	**
Salt		**	**	**	**	**
Plot site×S	alt	*	**	**	**	**

Different letters in each column show significant difference at $p \le 0.01$ (**) by Least Significant Difference (LSD). NS and * are represented non significant and significant at $p \le 0.05$, respectively.

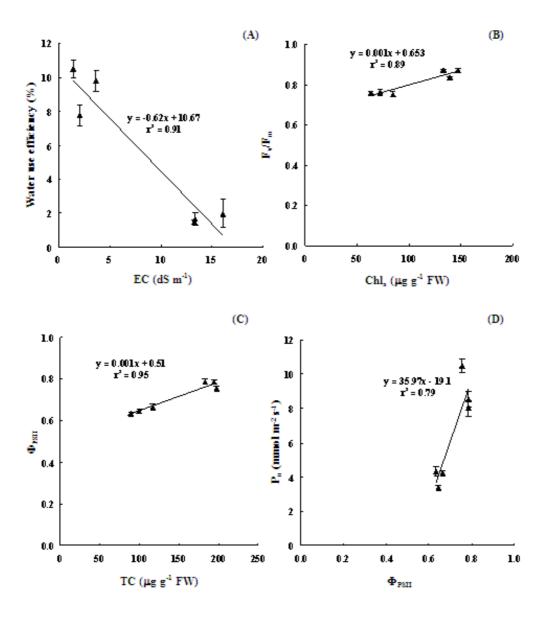


Figure 2. Relationships between electro conductivity in the soil and pigment content (A), chlorophyll a (Chl_a) and maximum quantum yield (F_v/F_m) (A), total chlorophyll (TC)and quantum efficiency of PSII (Φ_{PSII}) (B), quantum efficiency of PSII (Φ_{PSII}) and net photosynthetic rate (P_n) (C) in the leaf tissues of eucalypt tree cultivated in different salt-affected sites for 10 months.

The WUE in salt stressed eucalypt is significantly dropped, according to 100- 300 mol m^{-3} sodium salt in the pot culture (Adams et al., 2005). Reduction of WUE may be played as a major toxic to be induced pigment degradation, chlorophyll a fluorescence diminishing and net-photosynthetic rate (P_n) reduction when exposed to salt stress. Chlorophyll contents in E. camaldulensis cultivated in pot

culture containing salt are reduced (Rawat and Banerjee, 1998; Woodward and Bennett, 2005; Cha-um and Kirdmanee, 2008). Moreover, the chlorophyll a fluorescence activities of chlorophyll pigments, including maximum quantum yield of PSII (F_v/F_m) and quantum efficiency of PSII (Φ_{PSII}) are significantly diminished when exposed to salt-affected

soil (Cha-um and Kirdmanee, 2008), leading to reduce P_n with high g_s (Rawat and Banerjee, 1998) and may play as effective criteria to identify the salt adaptabilities in eucalypt species (Corney et al., 2003). The reduction percentages of photosynthetic abilities are positively

related to low biomass production i.e. plant height, diameter, number of leaves and plant dry weight (Rawat and Banerjee, 1998; Madsen and Mulligan, 2006; Chaum and Kirdmanee, 2008; Nasim et al., 2008).

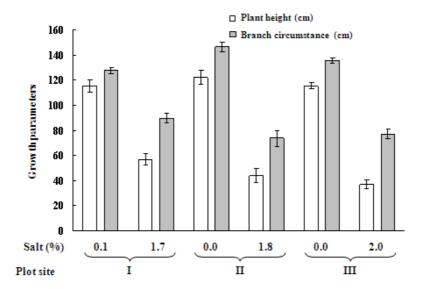


Figure 3. Plant height and circumstance diameter of eucalypt tree cultivated in different salt-affected sites for 10 months.

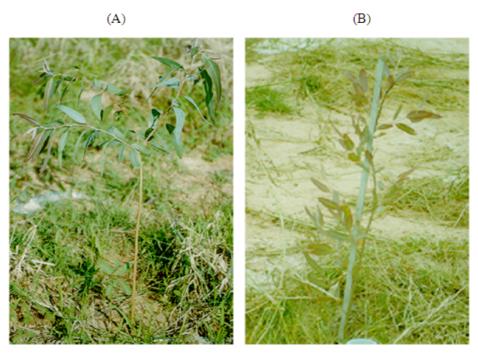


Figure 4. Morphological characters of eucalypt tree grown under control (0% salt) (A) and salt affected soil (2.0% salt) for 10 months.

In conclusion, the eucalypt plants can be grown under salt-affected soil containing <1.7% salt with > 30 survival percentage. The proline accumulation, water use efficiency, pigments and photosynthetic abilities as well as overall growth performance should be applied as indicator for salt tolerant screening of eucalypt tree. Eucalypt plant species may be further planted as a potential plant species to remediate the inland salt-affected soil.

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References

- Adams, M. A., A. Richter, A. K. Hill and T. D. Colmer. 2005. Salt tolerance in *Eucalyptus* spp.: identity and response of putative osmolytes. Plant Cell Environ. 28:772-787.
- Ahmad, R. and M. H. Chang. 2002. Salinity control and environmental protection through halophytes. J. Drain. Water Manage. 6:17-25.
- Bate, L. S., R. P. Waldren and I. D. Teare. 1973. Rapid determination of free proline for water-stress studies. Plant Soil 39:205-207.
- Cha-um, S. and C. Kirdmanee. 2008. Assessment of salt tolerance in *Eucalyptus*, rain tree and Thai neem under laboratory and the field conditions. Pak. J. Bot. 40:2041-2051.
- Cha-um, S., K. Mosaleeyanon, C. Kirdmanee and K. Supaibulwatana. 2003. A more efficient transplanting system for Thai neem (*Azadirachta siamensis* Val.) by reducing relative humidity. Sci. Asia 29:189-196.
- Cha-um, S., K. Supaibulwatana and C. Kirdmanee. 2007. Glycinebetaine accumulation, physiological characterizations, and growth efficiency

- in salt tolerant and salt sensitive lines of indica rice (*Oryza sativa* L. spp. *indica*) response to salt stress. J. Agron. Crops Sci. 193:157-166.
- Chinnusamy, V., A. Jagendorf and J. K. Zhu. 2005. Understanding and improving salt tolerance in plants. Crop Sci. 45:437-448.
- Corney, H. J., J. M. Sasse and P. K. Ades. 2003. Assessment of salt tolerance in eucalypts using chlorophyll fluorescence attributes. New Forest 26:233-246.
- Dale, G. and M. Dieters. 2007. Economic returns from environmental problems: Breeding salt- and drought-tolerant eucalypts for salinity abatement and commercial forestry. Ecol. Engineer. 31:175-182.
- Hasegawa, P. M., R. A. Bressan, J. K. Zhu and H. J. Bohnert. 2000. Plant cellular and molecular responses to high salinity. Annu. Rev. Plant Physiol. Plant Mol. Biol. 51:463-499.
- Jianfeng, Z., J. Jingmin and X. Shangjun. 2008. Planting techniques of *Tamarix chinensis* and its effect on saline soil remediation. IEEE doi: 978-1-4244-1748-3/08.
- Kiliç, C. C., Y. S. Kukul and D. Anaç. 2008. Performance of parslane (*Portulaca oleracea* L.) as a salt-removing crop. Agric. Water Manage. 95:854-858.
- Lichtenthaler, H. K. 1987. Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. Methods Enzymol. 148:350-380.
- Loggini, B., A. Scartazza, E. Brugnoli and F. Navari-Izzo. 1999. Antioxidant defense system, pigment composition, and photosynthetic efficiency in two wheat cultivars subjected to drought. Plant Physiol. 119:1091-1099.
- Maathuis, F. J. M. and A. Amtmann. 1999. K⁺ Nutrition and Na⁺ toxicity: The basis of cellular K⁺/Na⁺ ratios. Ann. Bot. 84:123-133.
- Madsen, P. A. and D. R. Mulligan. 2006. Effect of NaCl on emergence and growth

- of a range of provenances of *Eucalyptus ciriodora*, *Eucalyptus populnea*, *Eucalyptus camaldulensis* and *Acacia salicina*. Forest Ecol. Manage. 228:152-159.
- Mansour, M. M. F. and K. H. A. Salama. 2004. Cellular basis of salinity tolerance in plants. Env. Exp. Bot. 52:113-122.
- Nasim, M., R. Qureshi, T. Aziz, M. Saqib, S. Nawaz, S. T. Sahi and S. Pervaiz. 2008. Growth and ionic composition of salt-stressed *Eucalyptus camaldulensis* and *Eucalyptus tereticornis*. Pak. J. Bot. 40:799-805.
- Qadir, M., A. Tubeileh, J. Akhtar, A. Larbi, P. S. Minhas and M. A. Khan. 2008. Productivity enhancement of salt-affected environments through crop diversification. Land Degrad. Develop. 19:429-453.
- Qadir, M., J. D. Oster, S. Schubert, A. D. Noble and K. L. Sahrawat. 2007. Phytoremediation of sodic and saline-sodic soils. Adv. Agron. 96:197-247.
- Qadir, M. and J. D. Oster. 2004. Crop and irrigation management strategies for saline-sodic soils and water aimed at environmentally sustainable agriculture. Sci. Total Environ. 323:1-19.
- Rawat, J. S. and S. P. Banerjee. 1998. The influence of salinity on growth, biomass production and photosynthesis of *Eucalyptus camaldulensis* Dehnh. and *Dalbergia sissoo* Roxb. Seedlings. Plant Soil 205:163-169.
- Rockwood, D. L., A. W. Rudie, S. A. Ralph, J. Y. Zhu, and J. E. Winandy. 2008. Energy product options for *Eucalyptus* species grown as short rotation woody crops. Int. J. Mol. Sci. 9:1361-1378.
- Rodriguez-Navarro, A. and F. Rubio. 2006. High-affinity potassium and sodium transport systems in plants. J. Exp. Bot. 57:1149-1160.

- Shabala, S. N., S. I. Shabala, A.I. Martynenko, O. Babourina and I. A. Newman. 1998. Salinity effect on bioelectric activity, growth, Na⁺ accumulation and chlorophyll fluorescence of maize leaves: a comparative survey and prospects for screening. Aust. J. Plant Physiol. 25:609-616.
- Sharma, B. R. and P. S. Minhas. 2005. Strategies for managing saline/alkali waters for sustainable agricultural production in South Asia. Agric. Water Manage. 78:136-151.
- Shirazi, M. U., M. A. Khan, M. Ali, S. M. Mujtaba, S. Mumtaz, M. Ali, B. Khanzada, M. A. Halo, M. Rafique, J. A. Shah, K. A. Jafri and N. Depar. 2006. performance nutrient Growth and contents of some salt tolerant multipurpose tree species growing under saline environment. Pak. J. Bot. 38:1381-1388.
- Tanji, K. K. 2002. Salinity in the soil environment. In: A. Läuchli and U. Lüttge (Eds.), pp. 21-51. Salinity: Environment Plant Molecules, Kluwer Academic Publishers, Dordrecht, The Netherlands.
- Tester, M. and R. Davenport. 2003. Na⁺ tolerance and Na⁺ transport in higher plants. Ann. Bot. 91:503-527.
- Turner, N. C. and P. R. Ward. 2002. The role of agroforestry and perennial pasture in mitigating water logging and secondary salinity: summary. Agric. Water Manage. 53:271-275.
- Woodward, A. J. and I. J. Bennett. 2005. The effect of salt stress and abscisic acid on proline production, chlorophyll content and growth of *in vitro* propagated shoots of *Eucalyptus camaldulensis*. Plant Cell. Tiss. Org. Cult. 82:189-200.